

# Package: jackstraw (via r-universe)

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**Type** Package

**Title** Statistical Inference for Unsupervised Learning

**Version** 1.3.17

**Description** Test for association between the observed data and their estimated latent variables. The jackstraw package provides a resampling strategy and testing scheme to estimate statistical significance of association between the observed data and their latent variables. Depending on the data type and the analysis aim, the latent variables may be estimated by principal component analysis (PCA), factor analysis (FA), K-means clustering, and related unsupervised learning algorithms. The jackstraw methods learn over-fitting characteristics inherent in this circular analysis, where the observed data are used to estimate the latent variables and used again to test against that estimated latent variables. When latent variables are estimated by PCA, the jackstraw enables statistical testing for association between observed variables and latent variables, as estimated by low-dimensional principal components (PCs). This essentially leads to identifying variables that are significantly associated with PCs. Similarly, unsupervised clustering, such as K-means clustering, partition around medoids (PAM), and others, finds coherent groups in high-dimensional data. The jackstraw estimates statistical significance of cluster membership, by testing association between data and cluster centers. Clustering membership can be improved by using the resulting jackstraw p-values and posterior inclusion probabilities (PIPs), with an application to unsupervised evaluation of cell identities in single cell RNA-seq (scRNA-seq).

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find_k	<i>Find a number of clusters or principal components</i>
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### Description

There are a wide range of algorithms and visual techniques to identify a number of clusters or principal components embedded in the observed data.

### Usage

find\_k()

## Details

It is critical to explore the eigenvalues, cluster stability, and visualization. See R packages `bootcluster`, `EMCluster`, and `nFactors`.

Please see the R package `SC3`, which provides `estkTW()` function to find the number of significant eigenvalues according to the Tracy-Widom test.

`ADPclust` package includes `adpclust()` function that runs the algorithm on a range of  $K$  values. It helps you to identify the most suitable number of clusters.

This package also provides an alternative methods in `permutationPA`. Through a resampling-based Parallel Analysis, it finds a number of significant components.

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jackstraw

*jackstraw: Statistical Inference for Unsupervised Learning*

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## Description

Test for association between the observed data and their estimated latent variables. The `jackstraw` package provides a resampling strategy and testing scheme to estimate statistical significance of association between the observed data and their latent variables. Depending on the data type and the analysis aim, the latent variables may be estimated by principal component analysis (PCA), factor analysis (FA), K-means clustering, and related unsupervised learning algorithms. The `jackstraw` methods learn over-fitting characteristics inherent in this circular analysis, where the observed data are used to estimate the latent variables and used again to test against that estimated latent variables. When latent variables are estimated by PCA, the `jackstraw` enables statistical testing for association between observed variables and latent variables, as estimated by low-dimensional principal components (PCs). This essentially leads to identifying variables that are significantly associated with PCs. Similarly, unsupervised clustering, such as K-means clustering, partition around medoids (PAM), and others, finds coherent groups in high-dimensional data. The `jackstraw` estimates statistical significance of cluster membership, by testing association between data and cluster centers. Clustering membership can be improved by using the resulting `jackstraw` p-values and posterior inclusion probabilities (PIPs), with an application to unsupervised evaluation of cell identities in single cell RNA-seq (scRNA-seq).

## Details

The `jackstraw` package provides a resampling strategy and testing scheme to estimate statistical significance of association between the observed data and their latent variables. Depending on the data type and the analysis aim, the latent variables may be estimated by principal component analysis (PCA), K-means clustering, and related algorithms. The `jackstraw` methods learn over-fitting characteristics inherent in this circular analysis, where the observed data are used to estimate the latent variables and used again to test against those estimated latent variables.

The `jackstraw` tests enable us to identify the data features (i.e., variables or observations) that are driving systematic variation, in a completely unsupervised manner. Using `jackstraw_pca`, we can find statistically significant features with regard to the top  $r$  principal components. Alternatively, `jackstraw_kmeans` can identify the data features that are statistically significant members of the

data-dependent clusters. Furthermore, this package includes more general algorithms such as [jackstraw\\_subspace](#) for the dimension reduction techniques and [jackstraw\\_cluster](#) for the clustering algorithms.

Overall, it computes  $m$  p-values of association between the  $m$  data features and their corresponding latent variables. From  $m$  p-values, [pip](#) computes posterior inclusion probabilities, that are useful for feature selection and visualization.

### Author(s)

Neo Christopher Chung <nchchung@gmail.com>

### References

Chung and Storey (2015) Statistical significance of variables driving systematic variation in high-dimensional data. *Bioinformatics*, 31(4): 545-554 [doi:10.1093/bioinformatics/btu674](https://doi.org/10.1093/bioinformatics/btu674)

Chung (2020) Statistical significance of cluster membership for unsupervised evaluation of cell identities. *Bioinformatics*, 36(10): 3107–3114 [doi:10.1093/bioinformatics/btaa087](https://doi.org/10.1093/bioinformatics/btaa087)

### See Also

[jackstraw\\_pca](#) [jackstraw\\_subspace](#) [jackstraw\\_kmeans](#) [jackstraw\\_cluster](#)

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jackstraw\_alstructure *Non-Parametric Jackstraw for ALStructure*

---

### Description

Test association between the observed variables and population structure estimated by ALStructure.

### Usage

```
jackstraw_alstructure(  
  dat,  
  r,  
  FUN,  
  r1 = NULL,  
  s = NULL,  
  B = NULL,  
  covariate = NULL,  
  verbose = TRUE  
)
```

### Arguments

dat	a genotype matrix with m rows as variables and n columns as observations.
r	a number of significant LFs.
FUN	a function to ALStructure
r1	a numeric vector of LFs of interest (implying you are not interested in all r LFs).
s	a number of “synthetic” null variables. Out of m variables, s variables are independently permuted.
B	a number of resampling iterations. There will be a total of s*B null statistics.
covariate	a data matrix of covariates with corresponding n observations (do not include an intercept term).
verbose	a logical specifying to print the computational progress.

### Details

This function uses ALStructure from Cabrerros and Storey (2019). A deviation dev in logistic regression (the full model with r LFs vs. the intercept-only model) is used to assess association. This function also requires the Bioconductor gctest package to be installed.

### Value

jackstraw\_alstructure returns a list consisting of

p.value	m p-values of association tests between variables and their LFs
obs.stat	m observed deviances
null.stat	s*B null deviances

### Author(s)

Neo Christopher Chung <nchchung@gmail.com>

### References

Chung and Storey (2015) Statistical significance of variables driving systematic variation in high-dimensional data. *Bioinformatics*, 31(4): 545-554 [doi:10.1093/bioinformatics/btu674](https://doi.org/10.1093/bioinformatics/btu674)

### See Also

[jackstraw\\_pca](#) [jackstraw](#)

### Examples

```
## Not run:  
# load genotype data to analyze (not shown) into this variable  
X  
# choose the number of ancestries  
r <- 3
```

```

# load alstructure package (install from https://github.com/StoreyLab/alstructure)
library(alstructure)
# define the function this way, a function of the genotype matrix only
FUN <- function(x) t( alstructure(x, d_hat = r)$Q_hat )

# calculate p-values (and other statistics) for each SNP
out <- jackstraw_alstructure( X, r, FUN )

## End(Not run)

```

---

jackstraw\_cluster      *Jackstraw for the User-Defined Clustering Algorithm*

---

## Description

Test the cluster membership using a user-defined clustering algorithm

## Usage

```

jackstraw_cluster(
  dat,
  k,
  cluster,
  centers,
  algorithm = function(x, centers, ...) stats::kmeans(x, centers, ...),
  s = 1,
  B = 1000,
  center = TRUE,
  noise = NULL,
  covariate = NULL,
  pool = TRUE,
  verbose = FALSE,
  ...
)

```

## Arguments

dat	a data matrix with m rows as variables and n columns as observations.
k	a number of clusters.
cluster	a vector of cluster assignments.
centers	a matrix of all cluster centers.
algorithm	a clustering algorithm to use, where an output must include ‘cluster’ and ‘centers’. For exact specification, see <a href="#">kmeans</a> .
s	a number of “synthetic” null variables. Out of m variables, s variables are independently permuted.

B	a number of resampling iterations.
center	a logical specifying to center the rows. By default, TRUE.
noise	specify a parametric distribution to generate a noise term. If NULL, a non-parametric jackstraw test is performed.
covariate	a model matrix of covariates with n observations. Must include an intercept in the first column.
pool	a logical specifying to pool the null statistics across all clusters. By default, TRUE.
verbose	a logical specifying to print the computational progress. By default, FALSE.
...	additional, optional arguments to 'algorithm'.

### Details

The clustering algorithms assign  $m$  rows into  $K$  clusters. This function enable statistical evaluation if the cluster membership is correctly assigned. Each of  $m$  p-values refers to the statistical test of that row with regard to its assigned cluster. Its resampling strategy accounts for the over-fitting characteristics due to direct computation of clusters from the observed data and protects against an anti-conservative bias.

The user is expected to explore the data with a given clustering algorithm and determine the number of clusters  $k$ . Furthermore, provide `cluster` and `centers` as given by applying `algorithm` onto `dat`. The rows of `centers` correspond to  $k$  clusters, as well as available levels in `cluster`. This function allows you to specify a parametric distribution of a noise term. It is an experimental feature.

### Value

`jackstraw_cluster` returns a list consisting of

<code>F.obs</code>	$m$ observed F statistics between variables and cluster centers.
<code>F.null</code>	F null statistics between null variables and cluster centers, from the jackstraw method.
<code>p.F</code>	$m$ p-values of membership.

### Author(s)

Neo Christopher Chung <nchchung@gmail.com>

### References

Chung (2020) Statistical significance of cluster membership for unsupervised evaluation of cell identities. *Bioinformatics*, 36(10): 3107–3114 [doi:10.1093/bioinformatics/btaa087](https://doi.org/10.1093/bioinformatics/btaa087)

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jackstraw_irlba	<i>Non-Parametric Jackstraw for Principal Component Analysis (PCA) using the augmented implicitly restarted Lanczos bidiagonalization algorithm (IRLBA)</i>
-----------------	---

---

### Description

Test association between the observed variables and their latent variables captured by principal components (PCs). PCs are computed using the augmented implicitly restarted Lanczos bidiagonalization algorithm (IRLBA; see [irlba](#)).

### Usage

```
jackstraw_irlba(
  dat,
  r = NULL,
  r1 = NULL,
  s = NULL,
  B = NULL,
  covariate = NULL,
  verbose = TRUE,
  ...
)
```

### Arguments

dat	a data matrix with $m$ rows as variables and $n$ columns as observations.
r	a number (a positive integer) of significant principal components. See <a href="#">permutationPA</a> and other methods.
r1	a numeric vector of principal components of interest. Choose a subset of $r$ significant PCs to be used.
s	a number (a positive integer) of “synthetic” null variables. Out of $m$ variables, $s$ variables are independently permuted.
B	a number (a positive integer) of resampling iterations. There will be a total of $s*B$ null statistics.
covariate	a data matrix of covariates with corresponding $n$ observations (do not include an intercept term).
verbose	a logical specifying to print the computational progress.
...	additional arguments to <a href="#">irlba</a> .

### Details

This function computes  $m$  p-values of linear association between  $m$  variables and their PCs. Its resampling strategy accounts for the over-fitting characteristics due to direct computation of PCs from the observed data and protects against an anti-conservative bias.



Provide the data matrix, with  $m$  variables as rows and  $n$  observations as columns. Given that there are  $r$  significant PCs, this function tests for linear association between  $m$  variables and their  $r$  PCs.

You could specify a subset of significant PCs that you are interested in ( $r1$ ). If  $r1$  is given, then this function computes statistical significance of association between  $m$  variables and  $r1$ , while adjusting for other PCs (i.e., significant PCs that are not your interest). For example, if you want to identify variables associated with first and second PCs, when your data contains three significant PCs, set  $r=3$  and  $r1=c(1,2)$ .

Please take a careful look at your data and use appropriate graphical and statistical criteria to determine a number of significant PCs,  $r$ . The number of significant PCs depends on the data structure and the context. In a case when you fail to specify  $r$ , it will be estimated from a permutation test (Buja and Eyuboglu, 1992) using a function [permutationPA](#).

If  $s$  is not supplied,  $s$  is set to about 10% of  $m$  variables. If  $B$  is not supplied,  $B$  is set to  $m*10/s$ .

### Value

jackstraw\_irlba returns a list consisting of

p.value	$m$ p-values of association tests between variables and their principal components
obs.stat	$m$ observed F-test statistics
null.stat	$s*B$ null F-test statistics

### Author(s)

Neo Christopher Chung <nchchung@gmail.com>

### References

Chung and Storey (2015) Statistical significance of variables driving systematic variation in high-dimensional data. *Bioinformatics*, 31(4): 545-554 [doi:10.1093/bioinformatics/btu674](https://doi.org/10.1093/bioinformatics/btu674)

### See Also

[jackstraw](#) [jackstraw\\_subspace](#) [permutationPA](#)

### Examples

```
## simulate data from a latent variable model: Y = BL + E
B = c(rep(1,10),rep(-1,10), rep(0,180))
L = rnorm(20)
E = matrix(rnorm(200*20), nrow=200)
dat = B %>% t(L) + E
dat = t(scale(t(dat), center=TRUE, scale=TRUE))

## apply the jackstraw
out = jackstraw_irlba(dat, r=1)

## Use optional arguments
## For example, set s and B for a balance between speed of the algorithm and accuracy of p-values
## Not run:
```

```
## out = jackstraw_irlba(dat, r=1, s=10, B=200)
## End(Not run)
```

---

 jackstraw\_kmeans

*Non-Parametric Jackstraw for K-means Clustering*


---

## Description

Test the cluster membership for K-means clustering

## Usage

```
jackstraw_kmeans(
  dat,
  kmeans.dat,
  s = NULL,
  B = NULL,
  center = FALSE,
  covariate = NULL,
  match = TRUE,
  pool = TRUE,
  verbose = FALSE,
  ...
)
```

## Arguments

<code>dat</code>	a matrix with $m$ rows as variables and $n$ columns as observations.
<code>kmeans.dat</code>	an output from applying <code>kmeans()</code> onto <code>dat</code> .
<code>s</code>	a number of “synthetic” null variables. Out of $m$ variables, $s$ variables are independently permuted.
<code>B</code>	a number of resampling iterations.
<code>center</code>	a logical specifying to center the rows of the null samples. By default, TRUE.
<code>covariate</code>	a model matrix of covariates with $n$ observations. Must include an intercept in the first column.
<code>match</code>	a logical specifying to match the observed clusters and jackstraw clusters using minimum Euclidean distances.
<code>pool</code>	a logical specifying to pool the null statistics across all clusters. By default, TRUE.
<code>verbose</code>	a logical specifying to print the computational progress. By default, FALSE.
<code>...</code>	optional arguments to control the k-means clustering algorithm (refers to <code>kmeans</code> ).

## Details

K-means clustering assign  $m$  rows into  $K$  clusters. This function enable statistical evaluation if the cluster membership is correctly assigned. Each of  $m$  p-values refers to the statistical test of that row with regard to its assigned cluster. Its resampling strategy accounts for the over-fitting characteristics due to direct computation of clusters from the observed data and protects against an anti-conservative bias.

The input data (`dat`) must be of a class 'matrix'.

## Value

jackstraw\_kmeans returns a list consisting of

<code>F.obs</code>	$m$ observed F statistics between variables and cluster centers.
<code>F.null</code>	F null statistics between null variables and cluster centers, from the jackstraw method.
<code>p.F</code>	$m$ p-values of membership.

## Author(s)

Neo Christopher Chung <nchchung@gmail.com>

## References

Chung (2020) Statistical significance of cluster membership for unsupervised evaluation of cell identities. *Bioinformatics*, 36(10): 3107–3114 [doi:10.1093/bioinformatics/btaa087](https://doi.org/10.1093/bioinformatics/btaa087)

## Examples

```
## Not run:
dat = t(scale(t(Jurkat293T), center=TRUE, scale=FALSE))
kmeans.dat <- kmeans(dat, centers=2, nstart = 10, iter.max = 100)
jackstraw.out <- jackstraw_kmeans(dat, kmeans.dat)

## End(Not run)
```

---

jackstraw_kmeanspp	<i>Non-Parametric Jackstraw for K-means Clustering using RcppArmadillo</i>
--------------------	--

---

## Description

Test the cluster membership for K-means clustering, using K-means++ initialization

**Usage**

```
jackstraw_kmeanspp(
  dat,
  kmeans.dat,
  s = NULL,
  B = NULL,
  center = TRUE,
  covariate = NULL,
  verbose = FALSE,
  pool = TRUE,
  ...
)
```

**Arguments**

<code>dat</code>	a matrix with $m$ rows as variables and $n$ columns as observations.
<code>kmeans.dat</code>	an output from applying <code>ClusterR::KMeans_rcpp</code> onto <code>dat</code> .
<code>s</code>	a number of “synthetic” null variables. Out of $m$ variables, $s$ variables are independently permuted.
<code>B</code>	a number of resampling iterations.
<code>center</code>	a logical specifying to center the rows. By default, <code>TRUE</code> .
<code>covariate</code>	a model matrix of covariates with $n$ observations. Must include an intercept in the first column.
<code>verbose</code>	a logical specifying to print the computational progress. By default, <code>FALSE</code> .
<code>pool</code>	a logical specifying to pool the null statistics across all clusters. By default, <code>TRUE</code> .
<code>...</code>	optional arguments to control the k-means clustering algorithm (refers to <code>ClusterR::KMeans_rcpp</code> ).

**Details**

K-means clustering assign  $m$  rows into  $K$  clusters. This function enable statistical evaluation if the cluster membership is correctly assigned. Each of  $m$  p-values refers to the statistical test of that row with regard to its assigned cluster. Its resampling strategy accounts for the over-fitting characteristics due to direct computation of clusters from the observed data and protects against an anti-conservative bias.

Generally, it functions identical to `jackstraw_kmeans`, but this uses `ClusterR::KMeans_rcpp` instead of `stats::kmeans`. A speed improvement is gained by K-means++ initialization and `RcppArmadillo`. If the input data is still too large, consider using `jackstraw_MinibatchKmeans`.

The input data (`dat`) must be of a class ‘matrix’.

**Value**

`jackstraw_kmeanspp` returns a list consisting of

<code>F.obs</code>	$m$ observed F statistics between variables and cluster centers.
--------------------	--

F.null            F null statistics between null variables and cluster centers, from the jackstraw method.

p.F                m p-values of membership.

### Author(s)

Neo Christopher Chung <nchchung@gmail.com>

### References

Chung (2020) Statistical significance of cluster membership for unsupervised evaluation of cell identities. *Bioinformatics*, 36(10): 3107–3114 doi:[10.1093/bioinformatics/btaa087](https://doi.org/10.1093/bioinformatics/btaa087)

### Examples

```
## Not run:
library(ClusterR)
dat = t(scale(t(Jurkat293T), center=TRUE, scale=FALSE))
kmeans.dat <- KMeans_rcpp(dat, clusters = 10, num_init = 1,
max_iters = 100, initializer = 'kmeans++')
jackstraw.out <- jackstraw_kmeanspp(dat, kmeans.dat)

## End(Not run)
```

---

jackstraw\_lfa

*Non-Parametric Jackstraw for Logistic Factor Analysis*

---

### Description

Test association between the observed variables and their latent variables captured by logistic factors (LFs).

### Usage

```
jackstraw_lfa(
  dat,
  r,
  FUN,
  r1 = NULL,
  s = NULL,
  B = NULL,
  covariate = NULL,
  permute_alleles = TRUE,
  verbose = TRUE
)
```

**Arguments**

<code>dat</code>	either a genotype matrix with $m$ rows as variables and $n$ columns as observations, or a <code>BEDMatrix</code> object (see package <code>BEDMatrix</code> , these objects are transposed compared to the above but this works fine as-is, see example, no need to modify a <code>BEDMatrix</code> input). A <code>BEDMatrix</code> input triggers a low-memory mode where permuted data is also written and processed from disk, whereas a regular matrix input stores permutations in memory. The tradeoff is <code>BEDMatrix</code> version typically runs considerably slower, but enables analysis of very large data that is otherwise impossible.
<code>r</code>	a number of significant LFs.
<code>FUN</code>	a function to use for LFA.
<code>r1</code>	a numeric vector of LFs of interest (implying you are not interested in all $r$ LFs).
<code>s</code>	a number of “synthetic” null variables. Out of $m$ variables, $s$ variables are independently permuted.
<code>B</code>	a number of resampling iterations. There will be a total of $s*B$ null statistics.
<code>covariate</code>	a data matrix of covariates with corresponding $n$ observations (do not include an intercept term).
<code>permute_alleles</code>	If <code>TRUE</code> (default), alleles (rather than genotypes) are permuted, which results in a more Binomial synthetic null when data is highly structured. Changing to <code>FALSE</code> is not recommended, except for research purposes to confirm that it performs worse than the default.
<code>verbose</code>	a logical specifying to print the computational progress.

**Details**

This function uses logistic factor analysis (LFA) from Hao et al. (2016). Particularly, the deviance in logistic regression (the full model with  $r$  LFs vs. the intercept-only model) is used to assess significance. This function requires the `gctest` package, and in practice also the `lfa` package, to be installed from Bioconductor.

The random outputs of the regular matrix versus the `BEDMatrix` versions are equal in distribution. However, fixing a seed and providing the same data to both versions does not result in the same exact outputs. This is because the `BEDMatrix` version permutes loci in a different order by necessity.

**Value**

`jackstraw_lfa` returns a list consisting of

<code>p.value</code>	$m$ p-values of association tests between variables and their LFs
<code>obs.stat</code>	$m$ observed deviances
<code>null.stat</code>	$s*B$ null deviances

**Author(s)**

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## References

Chung and Storey (2015) Statistical significance of variables driving systematic variation in high-dimensional data. *Bioinformatics*, 31(4): 545-554 [doi:10.1093/bioinformatics/btu674](https://doi.org/10.1093/bioinformatics/btu674)

## See Also

[jackstraw\\_pca](#) [jackstraw](#) [jackstraw\\_subspace](#)

## Examples

```
## Not run:
## simulate genotype data from a logistic factor model: drawing rbinom from logit(BL)
m <- 5000; n <- 100; pi0 <- .9
m0 <- round(m*pi0)
m1 <- m - round(m*pi0)
B <- matrix(0, nrow=m, ncol=1)
B[1:m1,] <- matrix(runif(m1*n, min=-.5, max=.5), nrow=m1, ncol=n)
L <- matrix(rnorm(n), nrow=1, ncol=n)
BL <- B %*% L
prob <- exp(BL)/(1+exp(BL))

dat <- matrix(rbinom(m*n, 2, as.numeric(prob)), m, n)

# load lfa package (install from Bioconductor)
library(lfa)
# choose the number of logistic factors, including the intercept
r <- 2
# define the function this way, a function of the genotype matrix only
FUN <- function(x) lfa::lfa( x, r )

## apply the jackstraw_lfa
out <- jackstraw_lfa( dat, r, FUN )

# if you had very large genotype data in plink BED/BIM/FAM files,
# use BEDMatrix and save memory by reading from disk (at the expense of speed)
library(BEDMatrix)
dat_BM <- BEDMatrix( 'filepath' ) # assumes filepath.bed, .bim and .fam exist
# run jackstraw!
out <- jackstraw_lfa( dat_BM, r, FUN )

## End(Not run)
```

---

jackstraw\_MiniBatchKmeans

*Non-Parametric Jackstraw for Mini Batch K-means Clustering*

---

## Description

Test the cluster membership for K-means clustering

**Usage**

```
jackstraw_MiniBatchKmeans(
  dat,
  MiniBatchKmeans.output = NULL,
  s = NULL,
  B = NULL,
  center = TRUE,
  covariate = NULL,
  verbose = FALSE,
  batch_size = floor(nrow(dat)/100),
  initializer = "kmeans++",
  pool = TRUE,
  ...
)
```

**Arguments**

<code>dat</code>	a data matrix with $m$ rows as variables and $n$ columns as observations.
<code>MiniBatchKmeans.output</code>	an output from applying <code>ClusterR::MiniBatchKmeans()</code> onto <code>dat</code> . This provides more controls over the algorithm and subsequently the initial centroids used.
<code>s</code>	a number of “synthetic” null variables. Out of $m$ variables, $s$ variables are independently permuted.
<code>B</code>	a number of resampling iterations.
<code>center</code>	a logical specifying to center the rows. By default, <code>TRUE</code> .
<code>covariate</code>	a model matrix of covariates with $n$ observations. Must include an intercept in the first column.
<code>verbose</code>	a logical specifying to print the computational progress. By default, <code>FALSE</code> .
<code>batch_size</code>	the size of the mini batches.
<code>initializer</code>	the method of initialization. By default, <code>kmeans++</code> .
<code>pool</code>	a logical specifying to pool the null statistics across all clusters. By default, <code>TRUE</code> .
<code>...</code>	optional arguments to control the Mini Batch K-means clustering algorithm (refers to <code>ClusterR::MiniBatchKmeans</code> ).

**Details**

K-means clustering assign  $m$  rows into  $K$  clusters. This function enable statistical evaluation if the cluster membership is correctly assigned. Each of  $m$  p-values refers to the statistical test of that row with regard to its assigned cluster. Its resampling strategy accounts for the over-fitting characteristics due to direct computation of clusters from the observed data and protects against an anti-conservative bias.



**Value**

jackstraw\_MiniBatchKmeans returns a list consisting of

F.obs	m observed F statistics between variables and cluster centers.
F.null	F null statistics between null variables and cluster centers, from the jackstraw method.
p.F	m p-values of membership.

**Author(s)**

Neo Christopher Chung <nchchung@gmail.com>

**References**

Chung (2020) Statistical significance of cluster membership for unsupervised evaluation of cell identities. *Bioinformatics*, 36(10): 3107–3114 [doi:10.1093/bioinformatics/btaa087](https://doi.org/10.1093/bioinformatics/btaa087)

**Examples**

```
## Not run:
library(ClusterR)
dat = t(scale(t(Jurkat293T), center=TRUE, scale=FALSE))
MiniBatchKmeans.output <- MiniBatchKmeans(data=dat, clusters = 2, batch_size = 300,
initializer = "kmeans++")
jackstraw.output <- jackstraw_MiniBatchKmeans(dat,
MiniBatchKmeans.output = MiniBatchKmeans.output)

## End(Not run)
```

---

jackstraw\_pam

*Non-Parametric Jackstraw for Partitioning Around Medoids (PAM)*

---

**Description**

Test the cluster membership for Partitioning Around Medoids (PAM)

**Usage**

```
jackstraw_pam(
  dat,
  pam.dat,
  s = NULL,
  B = NULL,
  center = TRUE,
  covariate = NULL,
  verbose = FALSE,
  pool = TRUE,
  ...
)
```

**Arguments**

<code>dat</code>	a matrix with $m$ rows as variables and $n$ columns as observations.
<code>pam.dat</code>	an output from applying <code>cluster::pam()</code> on <code>dat</code> .
<code>s</code>	a number of “synthetic” null variables. Out of $m$ variables, $s$ variables are independently permuted.
<code>B</code>	a number of resampling iterations.
<code>center</code>	a logical specifying to center the rows. By default, TRUE.
<code>covariate</code>	a model matrix of covariates with $n$ observations. Must include an intercept in the first column.
<code>verbose</code>	a logical specifying to print the computational progress. By default, FALSE.
<code>pool</code>	a logical specifying to pool the null statistics across all clusters. By default, TRUE.
<code>...</code>	optional arguments to control the k-means clustering algorithm (refers to <code>kmeans</code> ).

**Details**

PAM assigns  $m$  rows into  $K$  clusters. This function enable statistical evaluation if the cluster membership is correctly assigned. Each of  $m$  p-values refers to the statistical test of that row with regard to its assigned cluster. Its resampling strategy accounts for the over-fitting characteristics due to direct computation of clusters from the observed data and protects against an anti-conservative bias.

For a large dataset, PAM could be too slow. Consider using `cluster::clara` and `jackstraw::jackstraw_clara`.

The input data (`dat`) must be of a class ‘matrix’.

**Value**

`jackstraw_pam` returns a list consisting of

<code>F.obs</code>	$m$ observed F statistics between variables and cluster medoids.
<code>F.null</code>	F null statistics between null variables and cluster medoids, from the <code>jackstraw</code> method.
<code>p.F</code>	$m$ p-values of membership.

**Author(s)**

Neo Christopher Chung <nchchung@gmail.com>

**References**

Chung (2020) Statistical significance of cluster membership for unsupervised evaluation of cell identities. *Bioinformatics*, 36(10): 3107–3114 [doi:10.1093/bioinformatics/btaa087](https://doi.org/10.1093/bioinformatics/btaa087)

## Examples

```
## Not run:
library(cluster)
dat = t(scale(t(Jurkat293T), center=TRUE, scale=FALSE))
pam.dat <- pam(dat, k=2)
jackstraw.out <- jackstraw_pam(dat, pam.dat = pam.dat)

## End(Not run)
```

---

jackstraw\_pca

*Non-Parametric Jackstraw for Principal Component Analysis (PCA)*

---

## Description

Test association between the observed variables and their latent variables captured by principal components (PCs).

## Usage

```
jackstraw_pca(
  dat,
  r = NULL,
  r1 = NULL,
  s = NULL,
  B = NULL,
  covariate = NULL,
  verbose = TRUE
)
```

## Arguments

dat	a data matrix with m rows as variables and n columns as observations.
r	a number (a positive integer) of significant principal components. See <a href="#">permutationPA</a> and other methods.
r1	a numeric vector of the principal components that are of interest. Choose a subset of r significant PCs to be used.
s	a number (a positive integer) of “synthetic” null variables. Out of m variables, s variables are independently permuted.
B	a number (a positive integer) of resampling iterations. There will be a total of s*B null statistics.
covariate	a data matrix of covariates with corresponding n observations (do not include an intercept term).
verbose	a logical specifying to print the computational progress.

## Details

This function computes  $m$  p-values of linear association between  $m$  variables and their PCs. Its resampling strategy accounts for the over-fitting characteristics due to direct computation of PCs from the observed data and protects against an anti-conservative bias.

Provide the data matrix, with  $m$  variables as rows and  $n$  observations as columns. Given that there are  $r$  significant PCs, this function tests for linear association between  $m$  variables and their  $r$  PCs.

You could specify a subset of significant PCs that you are interested in ( $r1$ ). If  $r1$  is given, then this function computes statistical significance of association between  $m$  variables and  $r1$ , while adjusting for other PCs (i.e., significant PCs that are not your interest). For example, if you want to identify variables associated with first and second PCs, when your data contains three significant PCs, set  $r=3$  and  $r1=c(1,2)$ .

Please take a careful look at your data and use appropriate graphical and statistical criteria to determine a number of significant PCs,  $r$ . The number of significant PCs depends on the data structure and the context. In a case when you fail to specify  $r$ , it will be estimated from a permutation test (Buja and Eyuboglu, 1992) using a function [permutationPA](#).

If  $s$  is not supplied,  $s$  is set to about 10% of  $m$  variables. If  $B$  is not supplied,  $B$  is set to  $m*10/s$ .

## Value

jackstraw\_pca returns a list consisting of

p.value	$m$ p-values of association tests between variables and their principal components
obs.stat	$m$ observed F-test statistics
null.stat	$s*B$ null F-test statistics

## Author(s)

Neo Christopher Chung <nchchung@gmail.com>

## References

Chung and Storey (2015) Statistical significance of variables driving systematic variation in high-dimensional data. *Bioinformatics*, 31(4): 545-554 [doi:10.1093/bioinformatics/btu674](https://doi.org/10.1093/bioinformatics/btu674)

## See Also

[jackstraw](#) [jackstraw\\_subspace](#) [permutationPA](#)

## Examples

```
## Not run:
## simulate data from a latent variable model: Y = BL + E
B = c(rep(1,50),rep(-1,50), rep(0,900))
L = rnorm(20)
E = matrix(rnorm(1000*20), nrow=1000)
dat = B %%% t(L) + E
dat = t(scale(t(dat), center=TRUE, scale=TRUE))
```

```
## apply the jackstraw
out = jackstraw_pca(dat, r=1)

## Use optional arguments
## For example, set s and B for a balance between speed of the algorithm and accuracy of p-values
## out = jackstraw_pca(dat, r=1, s=10, B=1000)

## End(Not run)
```

---

jackstraw_rpca	<i>Non-Parametric Jackstraw for Principal Component Analysis (PCA) using Randomized Singular Value Decomposition</i>
----------------	--

---

### Description

Test association between the observed variables and their latent variables captured by principal components (PCs). PCs are computed by randomized Singular Value Decomposition (see [rsvd](#)).

### Usage

```
jackstraw_rpca(
  dat,
  r = NULL,
  r1 = NULL,
  s = NULL,
  B = NULL,
  covariate = NULL,
  verbose = TRUE,
  ...
)
```

### Arguments

dat	a data matrix with m rows as variables and n columns as observations.
r	a number (a positive integer) of significant principal components. See <a href="#">permutationPA</a> and other methods.
r1	a numeric vector of principal components of interest. Choose a subset of r significant PCs to be used.
s	a number (a positive integer) of “synthetic” null variables. Out of m variables, s variables are independently permuted.
B	a number (a positive integer) of resampling iterations. There will be a total of s*B null statistics.
covariate	a data matrix of covariates with corresponding n observations (do not include an intercept term).
verbose	a logical specifying to print the computational progress.
...	additional arguments to rpca.

## Details

This function computes  $m$  p-values of linear association between  $m$  variables and their PCs. Its resampling strategy accounts for the over-fitting characteristics due to direct computation of PCs from the observed data and protects against an anti-conservative bias.

Provide the data matrix, with  $m$  variables as rows and  $n$  observations as columns. Given that there are  $r$  significant PCs, this function tests for linear association between  $m$  variables and their  $r$  PCs.

You could specify a subset of significant PCs that you are interested in ( $r1$ ). If  $r1$  is given, then this function computes statistical significance of association between  $m$  variables and  $r1$ , while adjusting for other PCs (i.e., significant PCs that are not your interest). For example, if you want to identify variables associated with first and second PCs, when your data contains three significant PCs, set  $r=3$  and  $r1=c(1,2)$ .

Please take a careful look at your data and use appropriate graphical and statistical criteria to determine a number of significant PCs,  $r$ . The number of significant PCs depends on the data structure and the context. In a case when you fail to specify  $r$ , it will be estimated from a permutation test (Buja and Eyuboglu, 1992) using a function [permutationPA](#).

If  $s$  is not supplied,  $s$  is set to about 10% of  $m$  variables. If  $B$  is not supplied,  $B$  is set to  $m*10/s$ .

## Value

jackstraw\_rpca returns a list consisting of

p.value	$m$ p-values of association tests between variables and their principal components
obs.stat	$m$ observed F-test statistics
null.stat	$s*B$ null F-test statistics

## Author(s)

Neo Christopher Chung <nchchung@gmail.com>

## References

Chung and Storey (2015) Statistical significance of variables driving systematic variation in high-dimensional data. *Bioinformatics*, 31(4): 545-554 [doi:10.1093/bioinformatics/btu674](https://doi.org/10.1093/bioinformatics/btu674)

## See Also

[jackstraw](#) [jackstraw\\_subspace](#) [permutationPA](#)

## Examples

```
## simulate data from a latent variable model: Y = BL + E
B = c(rep(1,10),rep(-1,10), rep(0,180))
L = rnorm(20)
E = matrix(rnorm(200*20), nrow=200)
dat = B %%% t(L) + E
dat = t(scale(t(dat), center=TRUE, scale=TRUE))

## apply the jackstraw
```

```

out = jackstraw_rpca(dat, r=1)

## Use optional arguments
## For example, set s and B for a balance between speed of the algorithm and accuracy of p-values
## Not run:
## out = jackstraw_rpca(dat, r=1, s=10, B=200)

## End(Not run)

```

---

jackstraw\_subspace      *Jackstraw for the User-Defined Dimension Reduction Methods*

---

## Description

Test association between the observed variables and their latent variables, captured by a user-defined dimension reduction method.

## Usage

```

jackstraw_subspace(
  dat,
  r,
  FUN,
  r1 = NULL,
  s = NULL,
  B = NULL,
  covariate = NULL,
  noise = NULL,
  verbose = TRUE
)

```

## Arguments

dat	a data matrix with $m$ rows as variables and $n$ columns as observations.
r	a number of significant latent variables.
FUN	Provide a specific function to estimate LVs. Must output $r$ estimated LVs in a $n \times r$ matrix.
r1	a numeric vector of latent variables of interest.
s	a number of “synthetic” null variables. Out of $m$ variables, $s$ variables are independently permuted.
B	a number of resampling iterations.
covariate	a model matrix of covariates with $n$ observations. Must include an intercept in the first column.
noise	specify a parametric distribution to generate a noise term. If NULL, a non-parametric jackstraw test is performed.
verbose	a logical specifying to print the computational progress.

## Details

This function computes  $m$  p-values of linear association between  $m$  variables and their latent variables, captured by a user-defined dimension reduction method. Its resampling strategy accounts for the over-fitting characteristics due to direct computation of PCs from the observed data and protects against an anti-conservative bias.

This function allows you to specify a parametric distribution of a noise term. It is an experimental feature. Then, a small number  $s$  of observed variables are replaced by synthetic null variables generated from a specified distribution.

## Value

jackstraw\_subspace returns a list consisting of

p.value	$m$ p-values of association tests between variables and their principal components
obs.stat	$m$ observed statistics
null.stat	$s \times B$ null statistics

## Author(s)

Neo Christopher Chung <nchchung@gmail.com>

## References

Chung and Storey (2015) Statistical significance of variables driving systematic variation in high-dimensional data. *Bioinformatics*, 31(4): 545-554 [doi:10.1093/bioinformatics/btu674](https://doi.org/10.1093/bioinformatics/btu674)

Chung (2020) Statistical significance of cluster membership for unsupervised evaluation of cell identities. *Bioinformatics*, 36(10): 3107–3114 [doi:10.1093/bioinformatics/btaa087](https://doi.org/10.1093/bioinformatics/btaa087)

## See Also

[jackstraw\\_pca](#) [jackstraw](#)

## Examples

```
## simulate data from a latent variable model: Y = BL + E
B = c(rep(1,50),rep(-1,50), rep(0,900))
L = rnorm(20)
E = matrix(rnorm(1000*20), nrow=1000)
dat = B %>% t(L) + E
dat = t(scale(t(dat), center=TRUE, scale=TRUE))

## apply the jackstraw with the svd as a function
out = jackstraw_subspace(dat, FUN = function(x) svd(x)$v[,1,drop=FALSE], r=1, s=100, B=50)
```



---

Jurkat293T

*A Jurkat:293T equal mixture dataset from Zheng et al. (2017)*

---

**Description**

50

**Usage**

Jurkat293T

**Format**

A data frame with 3381 rows corresponding to single cells and 10 columns corresponding to the top 10 principal components

**Source**

Supplementary Data 1 from Zheng et al. (2017) [https://static-content.springer.com/esm/art%3A10.1038%2Fncomms14049/MediaObjects/41467\\_2017\\_BFncomms14049\\_MOESM829\\_ESM.xlsx](https://static-content.springer.com/esm/art%3A10.1038%2Fncomms14049/MediaObjects/41467_2017_BFncomms14049_MOESM829_ESM.xlsx)

**References**

Zheng et al. (2017) Massively parallel digital transcriptional profiling of single cells. Nature Communications. 8:14049. doi:10.1038/ncomms14049

---

permutationPA

*Permutation Parallel Analysis*

---

**Description**

Estimate a number of significant principal components from a permutation test.

**Usage**

```
permutationPA(dat, B = 100, threshold = 0.05, verbose = TRUE)
```

**Arguments**

dat	a data matrix with m rows as variables and n columns as observations.
B	a number (a positive integer) of resampling iterations.
threshold	a numeric value between 0 and 1 to threshold p-values.
verbose	a logical indicator as to whether to print the progress.

**Details**

Adopted from `sva::num.sv`, and based on Buja and Eyuboglu (1992)

**Value**

`permutationPA` returns

`r` an estimated number of significant principal components based on thresholding p-values at threshold

`p` a list of p-values for significance of principal components

**References**

Buja A and Eyuboglu N. (1992) Remarks on parallel analysis. *Multivariate Behavioral Research*, 27(4), 509-540

---

`pip` *Compute posterior inclusion probabilities (PIPs)*

---

**Description**

From a set of p-values, computes posterior probabilities that a feature should be truly included. For example, membership inclusion in a given cluster can be improved by filtering low quality members. In using PCA and related methods, it helps select variables that are truly associated with given latent variables.

**Usage**

```
pip(pvalue, group = NULL, pi0 = NULL, verbose = TRUE, ...)
```

**Arguments**

`pvalue` a vector of p-values.

`group` a vector of group indicators (optional). If provided, PIP analysis is stratified. Assumes groups are in 1:k where k is the number of unique groups.

`pi0` a vector of pi0 values (optional). Its length has to be either 1 or equal the number of groups.

`verbose` If TRUE, reports information.

`...` optional arguments for `lfdr` to control a local FDR estimation.

**Details**

This function requires the Bioconductor `qvalue` package to be installed.

**Value**

`pip` returns a vector of posterior inclusion probabilities

**Author(s)**

Neo Christopher Chung <nchchung@gmail.com> John R. Yamamoto-Wilson

**References**

Chung (2020) Statistical significance of cluster membership for unsupervised evaluation of cell identities. *Bioinformatics*, 36(10): 3107–3114 doi:[10.1093/bioinformatics/btaa087](https://doi.org/10.1093/bioinformatics/btaa087)

Chung (2014) "Jackstraw Weighted Shrinkage for Principal Component Analysis and Covariance Matrix" in *Statistical Inference of Variables Driving Systematic Variation in High-Dimensional Biological Data*. PhD thesis, Princeton University. <https://www.proquest.com/openview/e90b562d689cf3a021c35a93c6f31/pq-origsite=gscholar&cbl=18750>

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